

Reducing psyllids improves tree health even when trees have HLB: IPM approaches

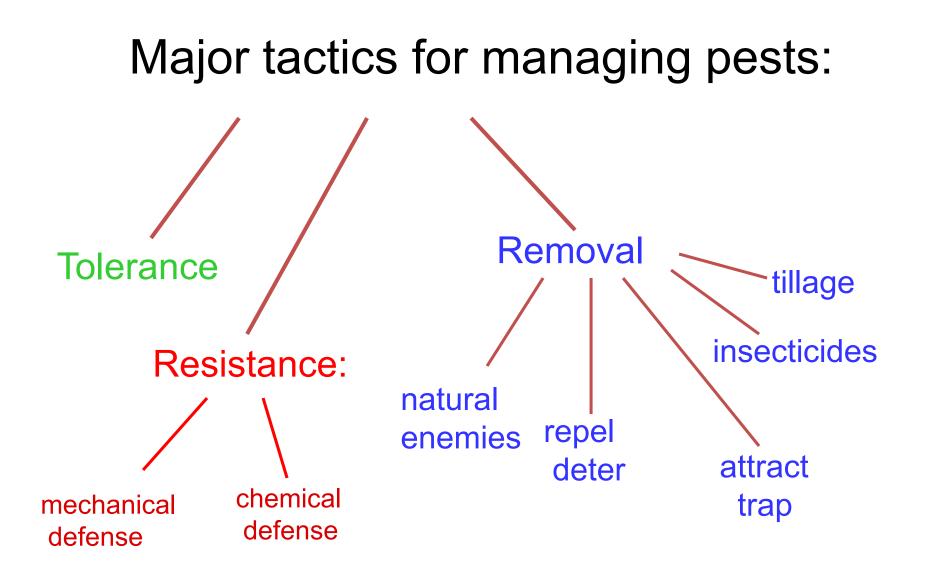
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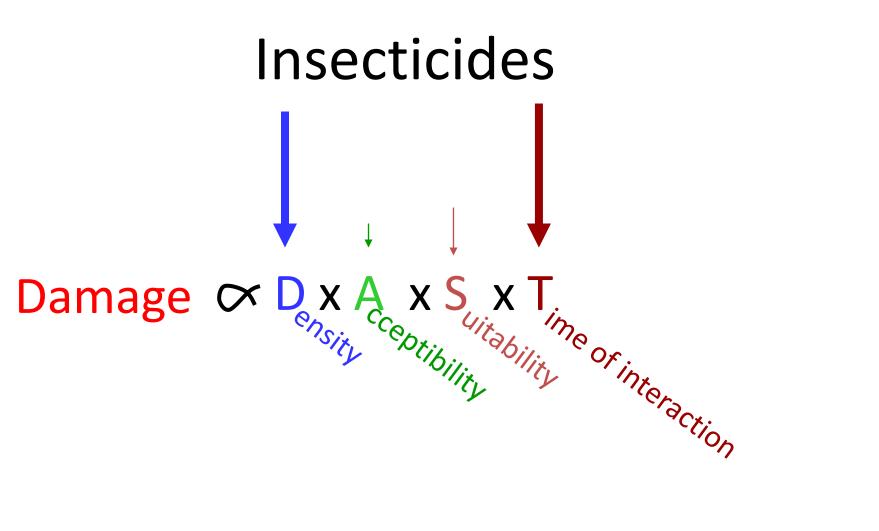


Parameters Damage C D x A x S x T ensity of ceptibility of crop of interaction







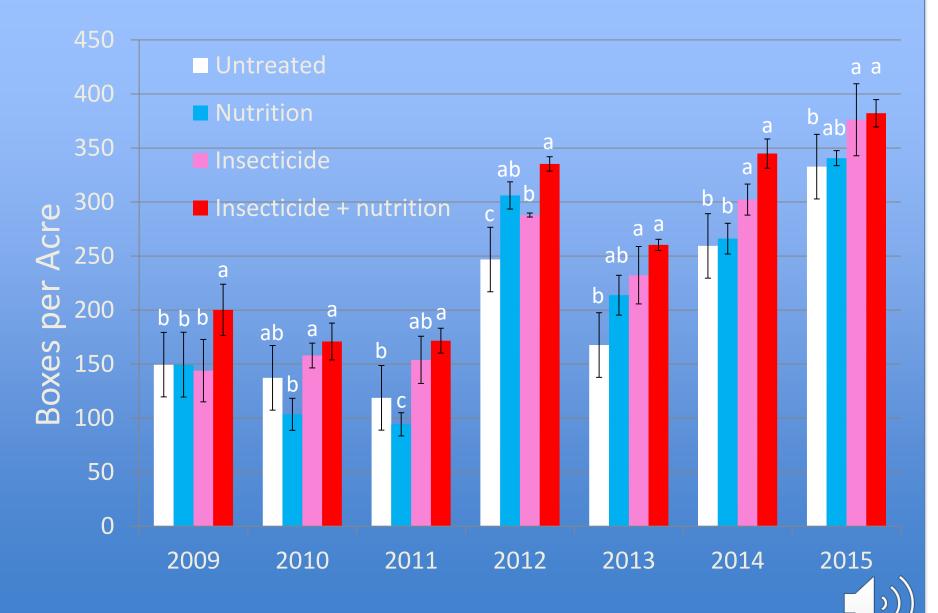




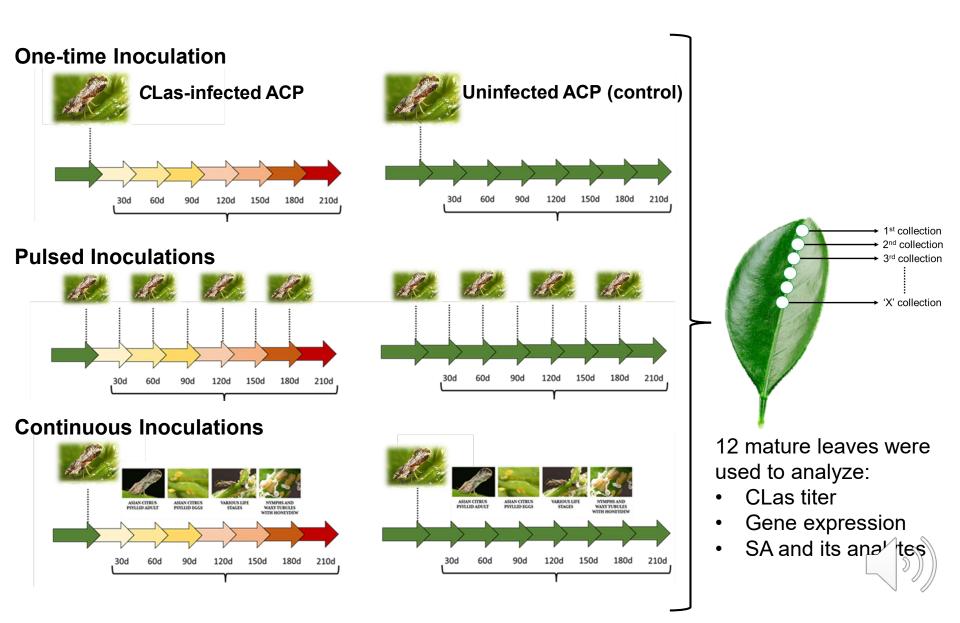
Resistant Varieties Damage C D x A x S x T, *b* x A x S x T, *c*ceptibility of interaction



Keeping ACP down seems to help yield



Experimental design



Pathogen titer not related to inoculation frequency

CLas titer



- No differences in CLas titer between plants exposed to infected vectors once (7 day inoculation access period) as compared with plants exposed to continuously to breeding CLas-infected insects
- Are insecticide applications needed if CLas titer does not appear to be impacted (no super-infection)?

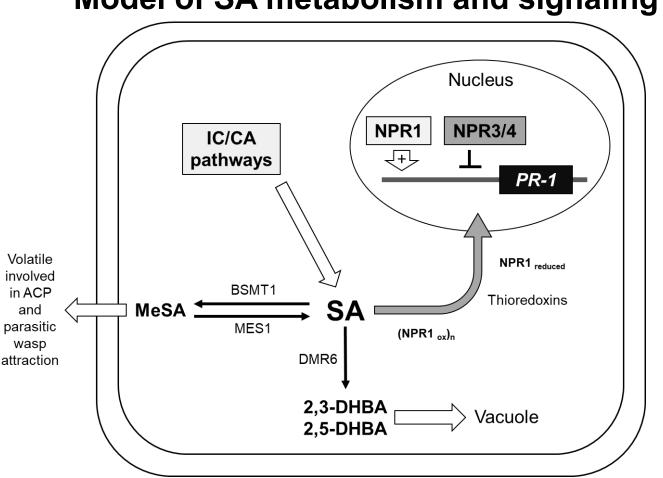
Larger cage experiment in Texas (Mamoudou Setamou, Texas A&M)-same results

HLB +; No ACP	HLB +; Pulsed (mont	hly) ACP	HLB +; Continuous ACP
		PI	ant health
		CLas- infected ACP	
HLB -; No ACP	HLB -; Pulsed (mo		ate Good _P
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Do CLas and ACP affect SA defenses?

We have described salicylic acid-induced plant defense response at varying

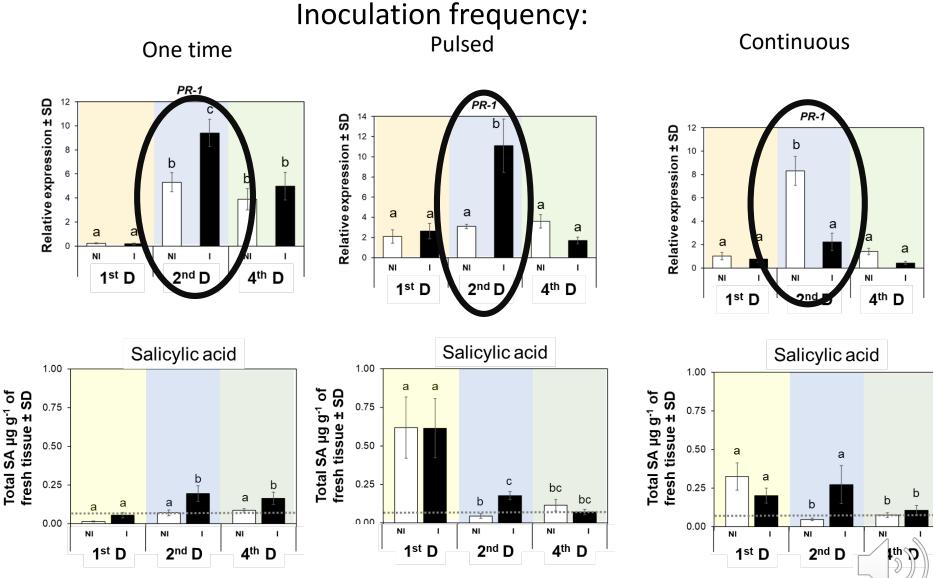
CLas inoculation frequencies



Model of SA metabolism and signaling

ACP damage interacts with CLas to affect plant defense:

White bars = Uninfected Black bars = CLas-infected



Take home

- Pulses of psyllid feeding stimulate immunity [increase salicylic acid (SA)], but long-term feeding turns it off [decreases SA and increases the genes (*BMST* and *DMR6*) that metabolize SA].
- After one-time inoculation, CLas-infected samples exhibit upregulation of *PR1* genes; these turn on defense response after a long period of infection (270 and 330 days).
- CLas-infected trees without other stressors (psyllid damage) can induce an immune response against CLas infection by activating SA-dependent defense responses.

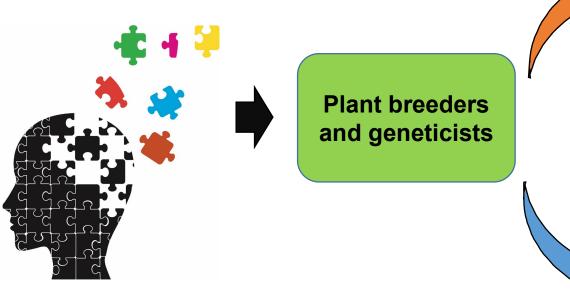


Take home

- Psyllid feeding also increases NPR3/4 gene expression.
- NPR3/4 reduces expression of genes involved in plant defenses (PR1) when SA concentrations are low.
- CLas infection with ACP damage causes a situation where defense response is compromised.
- The elimination of ACP from the tritrophic interaction allows the citrus plant to better handle CLas infection.

Future directions

- We hypothesize that enhanced resistance to CLas will be observed in transgenic plants with low expression of NPR3/NPR4
- We propose to determine the biological functions on NPR3/NPR4 in HLB disease progression and describe their role in SA signaling. To accomplish this goal, CLas-susceptible 'Hamlin', and CLastolerant 'Bearss' lemon will be transformed for overexpression/knockdown expression of NPR3/NPR4





Downregulation

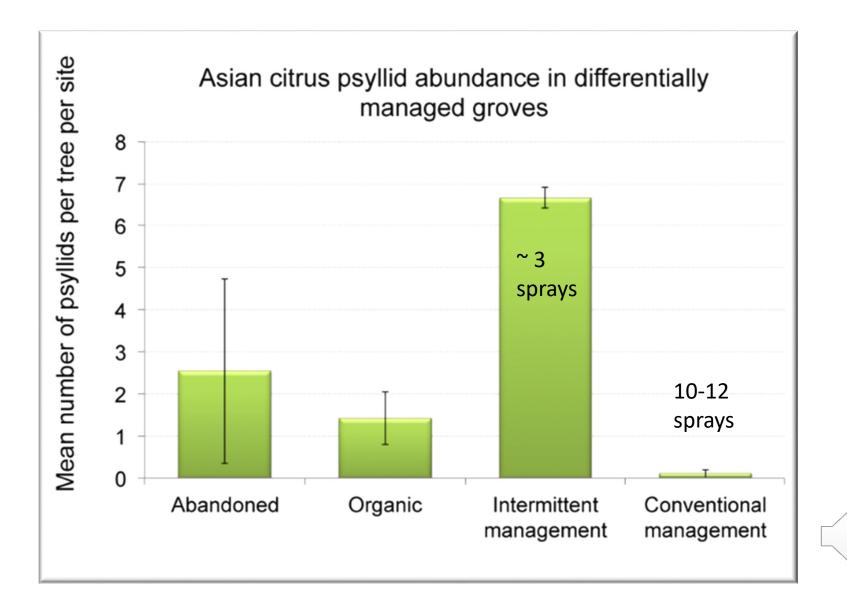
Upregulation

General observations

- CLas titer in leaf tissues is not affected by inoculum load imparted by the vector
- Plants respond to pulses of ACP feeding with boost to plant defense
- Long-term ACP feeding suppresses plant immunity and inhibits growth, which explains the importance of vector suppression as part of HLB management
- ACP management is beneficial, but we need to make it more sustainable.



Psyllids can be pushed down with intense use of insecticides, but this is not sustainable for many (2016-17) – Average from 4-13 groves



Typical recent model for ACP sprays:

- After harvest, a dormant spray has been usually timed before major spring flush using pyrethroid or organophosphate.
- Sprays made on a schedule with intervals somewhat determined by length of efficacy of a particular insecticide.

Possible better alternative:

- Spray for adults at bud break at the beginning of each new flush before there is feather flush on which adults can lay eggs.
- Apply second spray on the flush as ACP begin to reappear. This seems to achieve more than 60 days of low ACP populations
- Hold off spraying until ACP reach threshold (0.2—0.7 per tap)

Objectives

 Evaluate conventional grower practices versus sprays coordinated with bud break to suppress ACP populations

Specific Objectives

- a) Quantify impact of management protocols on ACP populations and CLas titer.
- b) Examine association between insecticide applications, ACP populations and CLas titer.



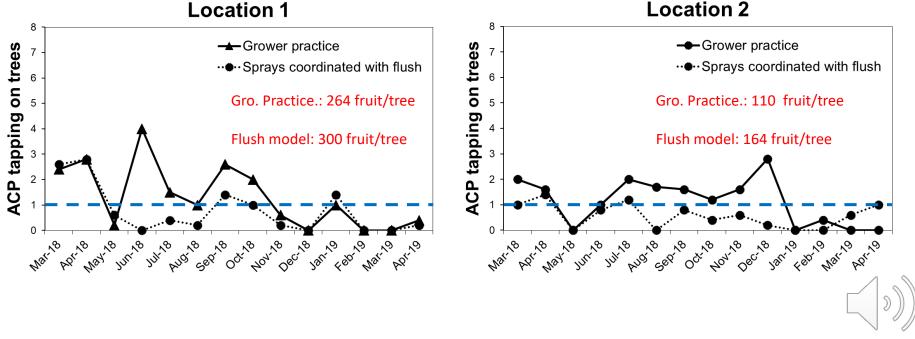
Take Home

- Timing insecticide applications with bud break caused better ACP suppression.
- Maintaining ACP populations below a threshold of 1 ACP / tap was associated with better yield.

-Solid lines indicate applications near flushing periods-grower practice (higher ACP)

-Dashed lines indicate use of bud break model to predict flushes (lower ACP)

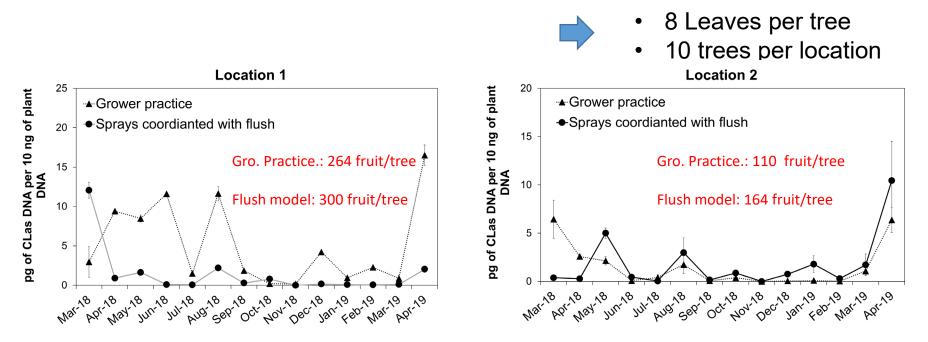
Location 2



Take Home

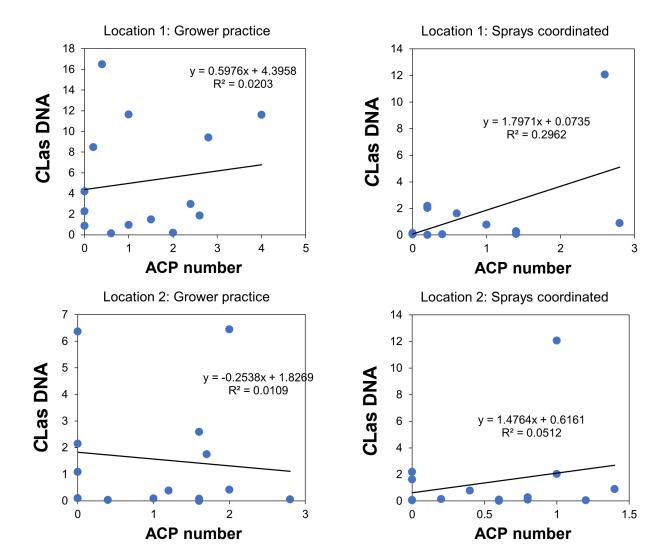
CLas titer over time.

Higher fruit-yields were associated with lower ACP populations when sprays were coordinated with flushing cycles using bud break phenology model

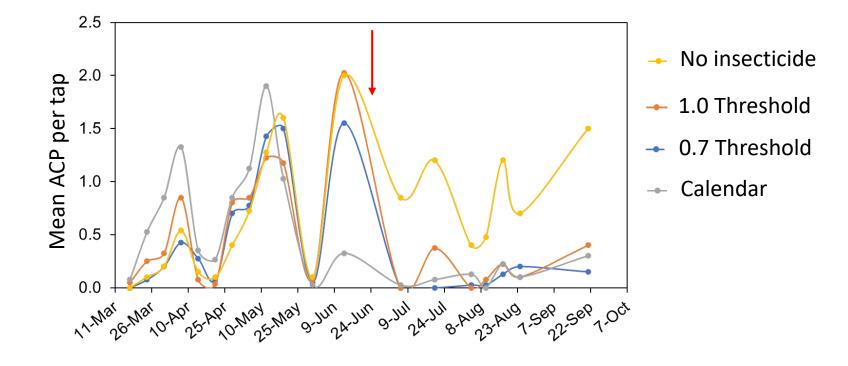


Take home

No correlation between ACP density and CLas titer—*This field result is congruent with our growth chamber investigations.* The difference in yield was more likely caused by difference in ACP damage



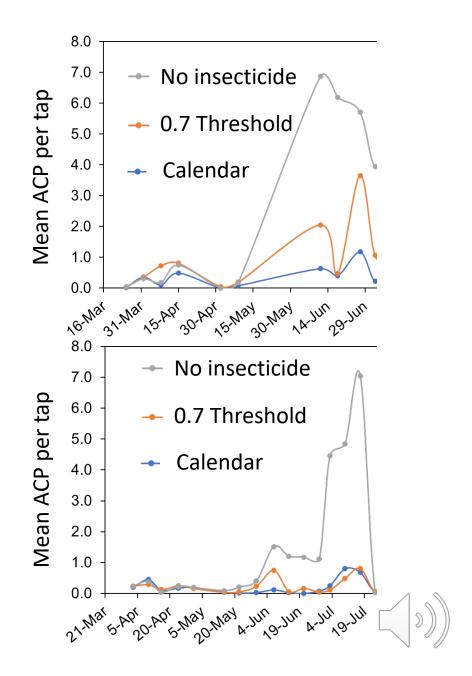
Psyllid densities when using thresholds were slightly higher than with calendar sprays





Key Feature of IPM:

- Deciding whether treatment is necessary after assessing the pest populations
- This decisions based on regular monitoring
- You determine whether to spray using a threshold that tells you that you reached a point where there are "too many psyllids" present rather than spraying on a calendar basis
- In our case, it's the point where cumulative annual damage reduces yield
- 0.7 ACP/tap looks like a pretty good ballpark threshold



Take home

- CLas titer in trees does not appear related to psyllid population density
- Psyllid density is related to tree stress—more psyllids--> higher damage (stress), which compromises tree health
- If the pest population (and the resulting damage) is low enough, it does not pay to take control measures
- As the pest population continues to rise, it reaches a point where the resulting damage would justify taking control measures
- 0.7 ACP / tap is a "working ballpark" threshold

