

Antimicrobial strategies for controlling CLas and the Asian citrus psyllid

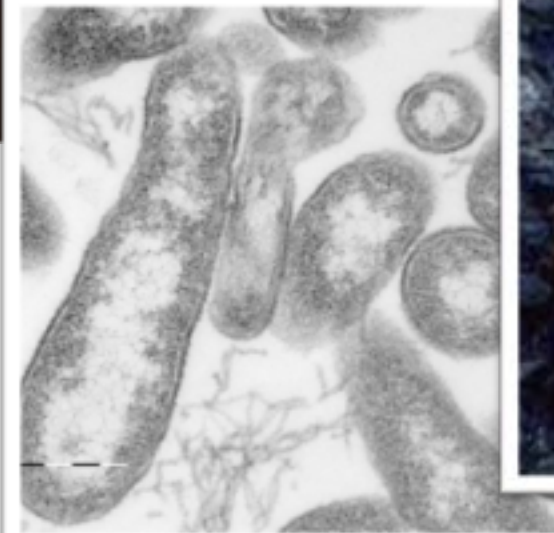
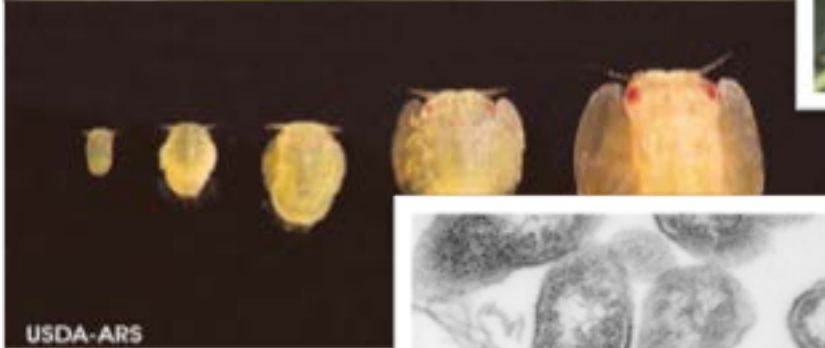
Kirsten Pelz-Stelinski

Associate Professor

Citrus Research and Education Center



Citrus greening disease or Huanglongbing



Current psyllid management requires significant input of insecticides

- Use of broad-spectrum insecticides targeting adult psyllids with possible rotation with insect growth regulators
- Applications made prior to new flush are most effective in reducing psyllid populations
 - 10-12+ annual sprays annually



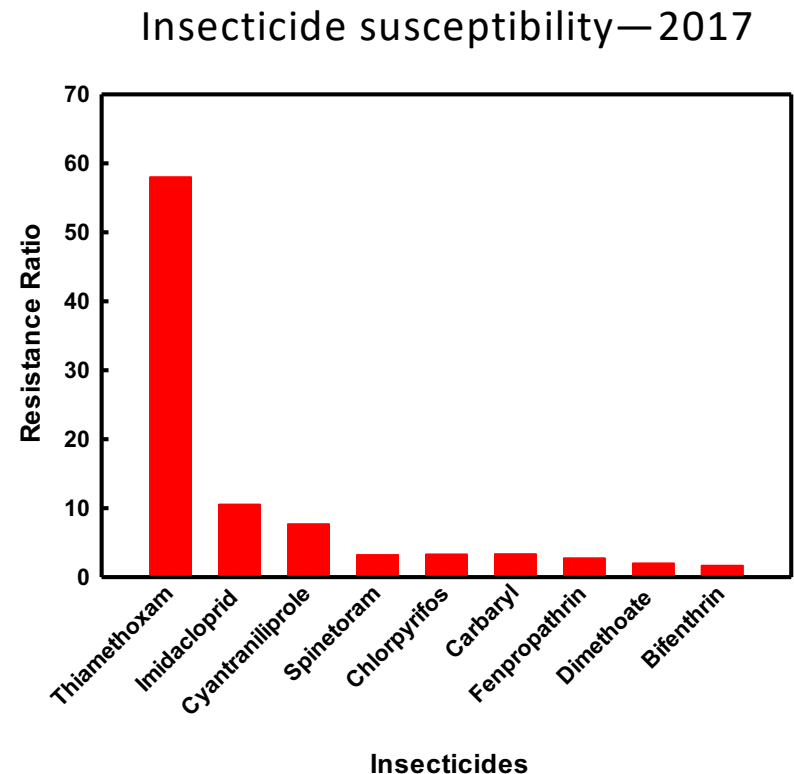
Antimicrobials for reducing pathogen transmission

Current management of psyllids with broad spectrum insecticides is unsustainable

- High cost
- Physiological resistance

Challenge: Targeted manipulation of symbionts provides a unique opportunity for vector/pathogen management

- Symbiont elimination/suppression
 - Antibiotics
 - Bactericides: oligonucleotide suppression of Las and endosymbionts



Antimicrobials for reducing pathogen transmission

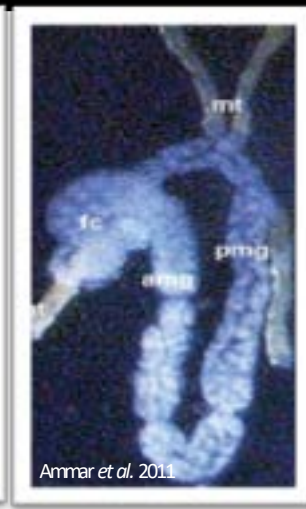
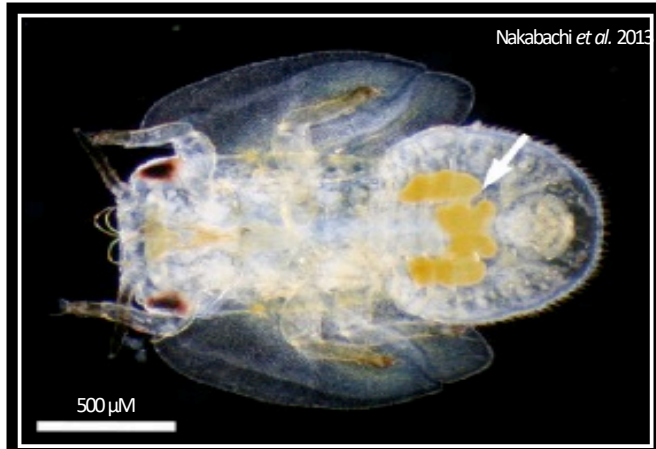
Current management of psyllids with broad spectrum insecticides is unsustainable

- High cost
- Physiological resistance

Can we manipulate microorganisms present in the vector and host plant to reduce pathogen transmission?

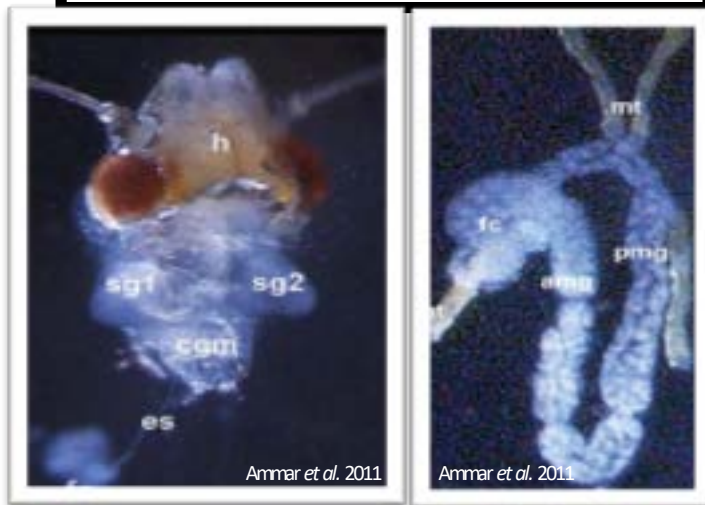
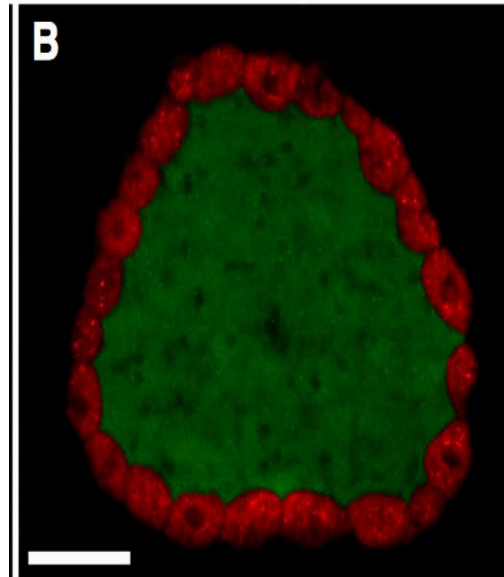
- Disruption of symbionts for ACP/CLas management

ACP bacterial endosymbionts



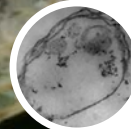
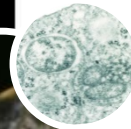
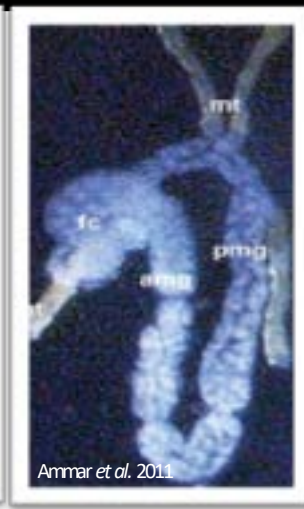
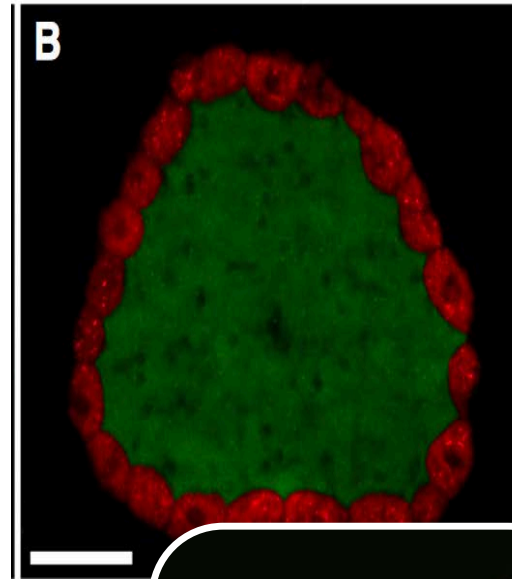
- *Candidatus Carsonella ruddii*
- *Candidatus Profftella armature*
- *Wolbachia* (wDi)
- *Candidatus Liberibacter asiaticus*
- *Wolbachia* (wDi)

ACP bacterial endosymbionts

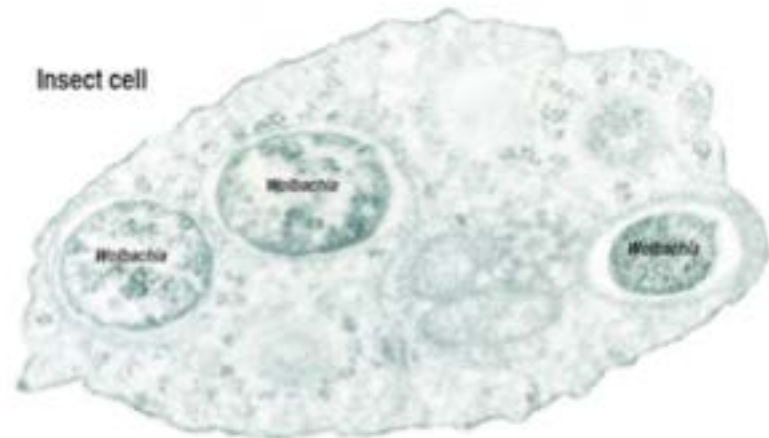
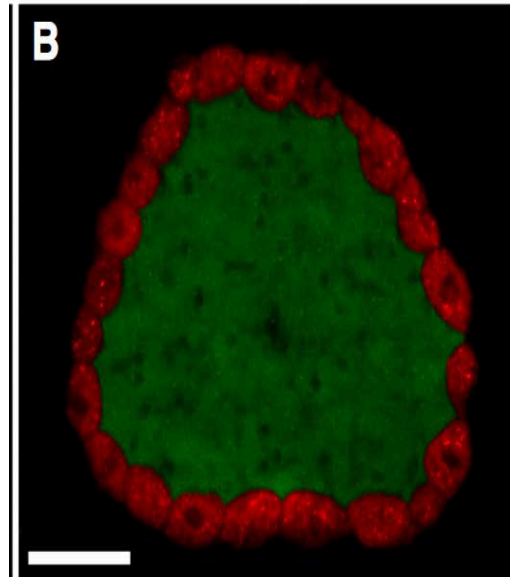
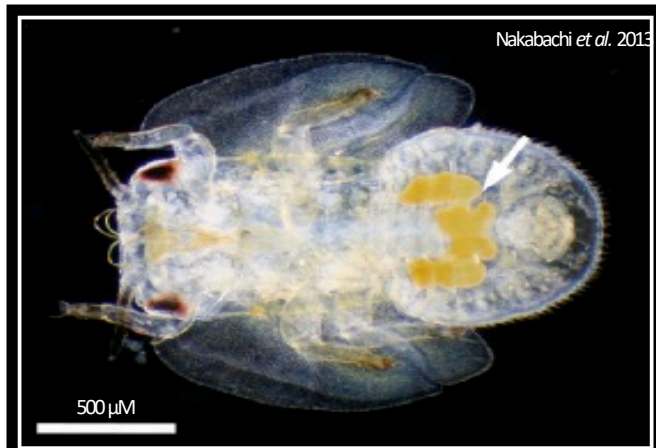


- *Candidatus Liberibacter asiaticus*
- *Wolbachia* (wDi)

ACP bacterial endosymbionts



ACP bacterial endosymbionts



Antimicrobials for reducing ACP survival and Las transmission

Antibiotic treatments:

- Plants (Las)
- ACP (Las, endosymbionts)

Antisense RNA technologies:

- Reduce Las in plants
- Selectively eliminate Las symbionts

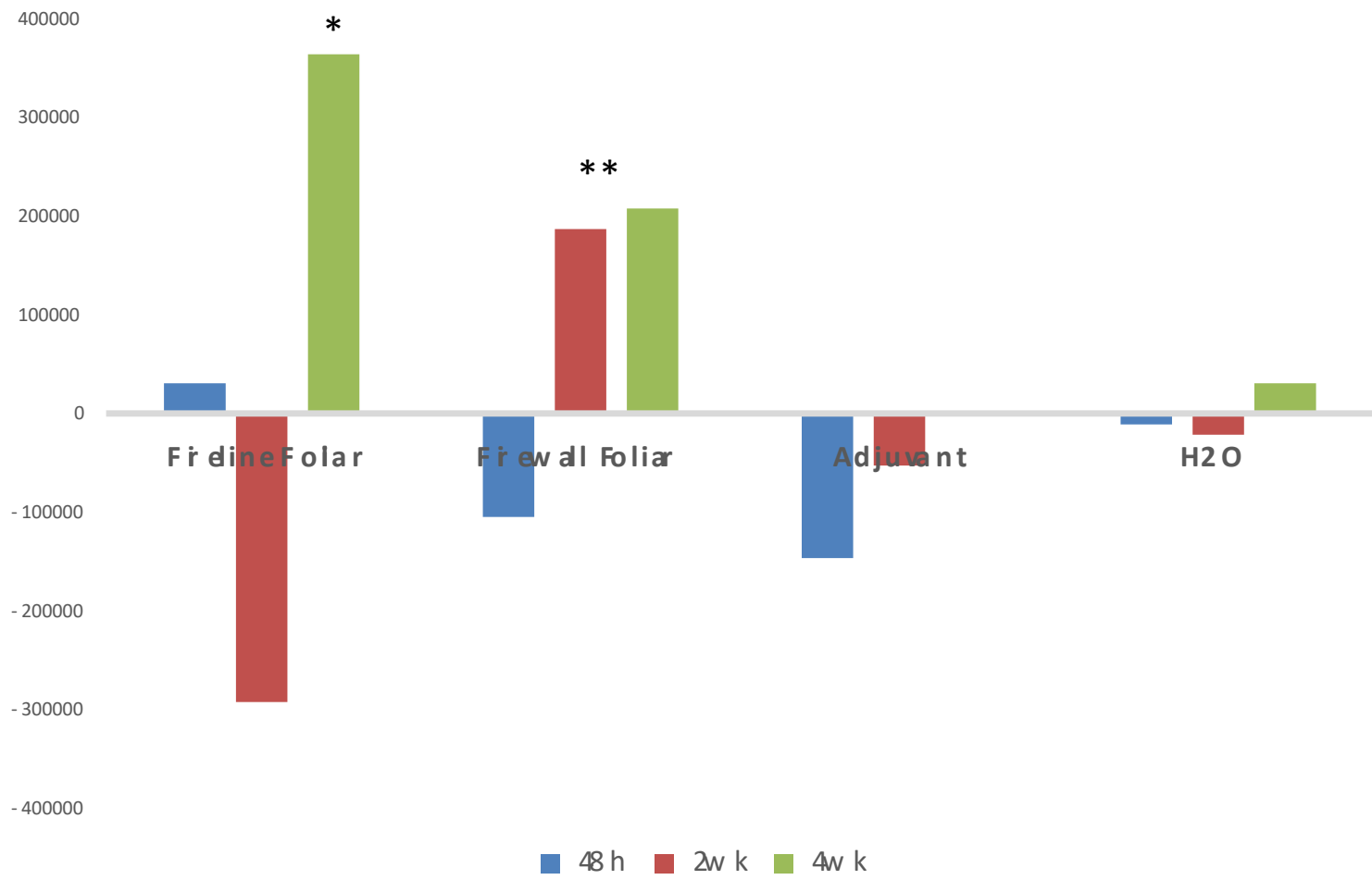
Management of Huanglongbing: EPA crisis declaration

Florida citrus lost an estimated \$7.80 billion between 2006-2014.

Three commercially available bactericides for Florida citrus:

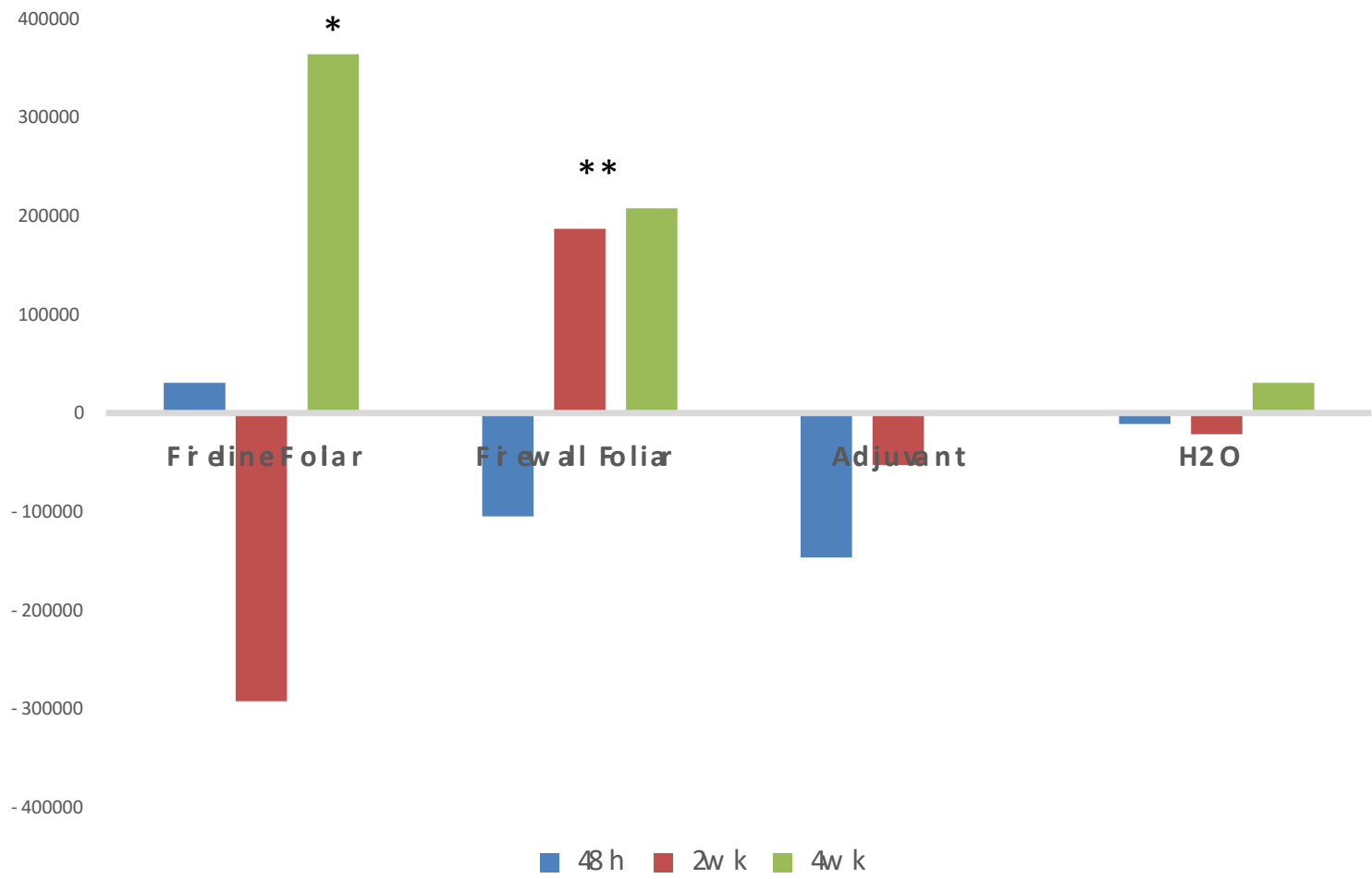
- FireWall™ 50 WP: Streptomycin sulfate
- FireLine™ 17 WP: Oxytetracycline hydrochloride
- Mycoshield®: Oxytetracycline calcium complex base

Greenhouse Plant Assay: Firewall Vs. Fireline Foliar Spray Efficacy



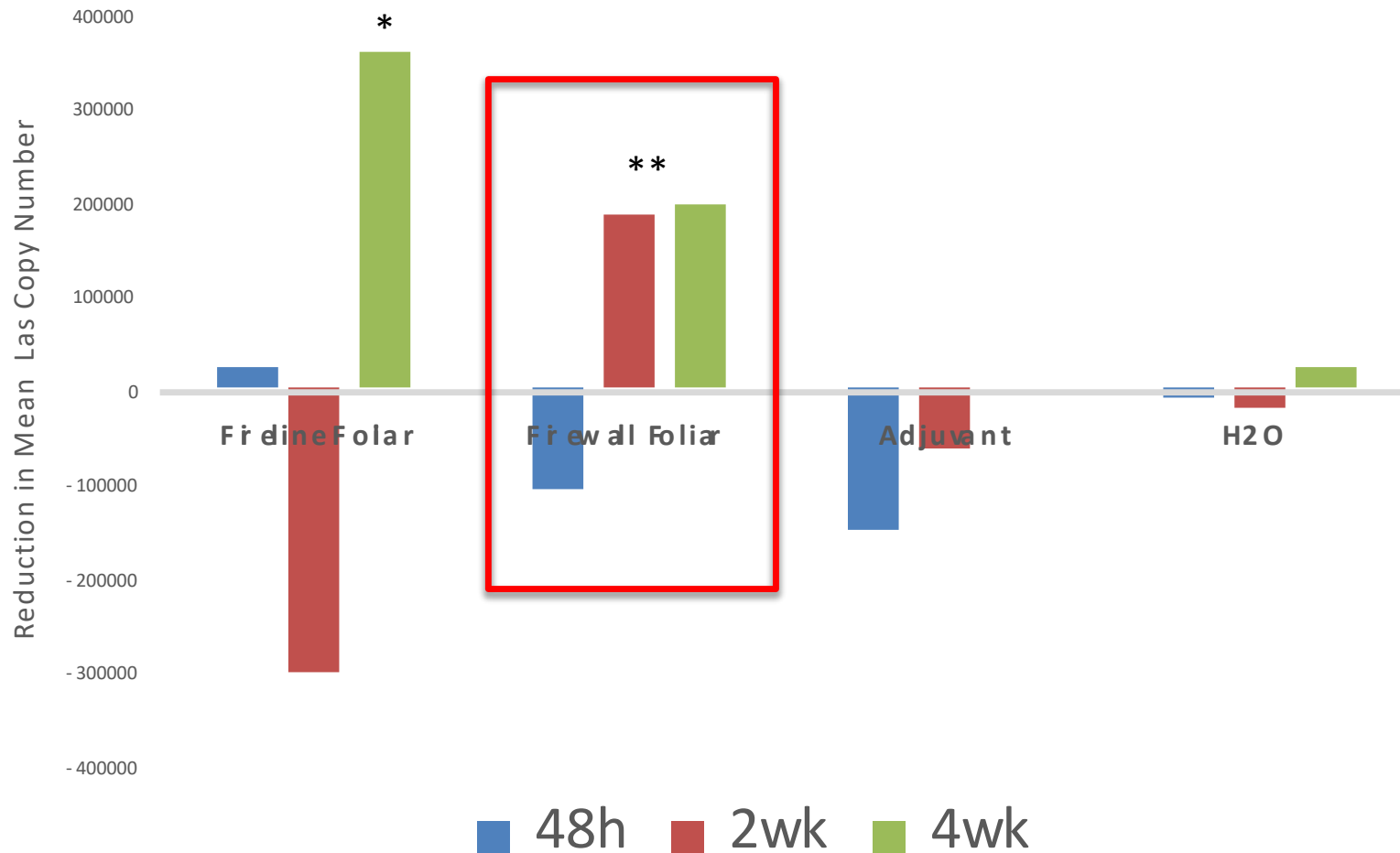
FireWall and Fireline applied at label rate (200 PPM)

Greenhouse Plant Assay: Firewall Vs. Fireline Foliar Spray Efficacy



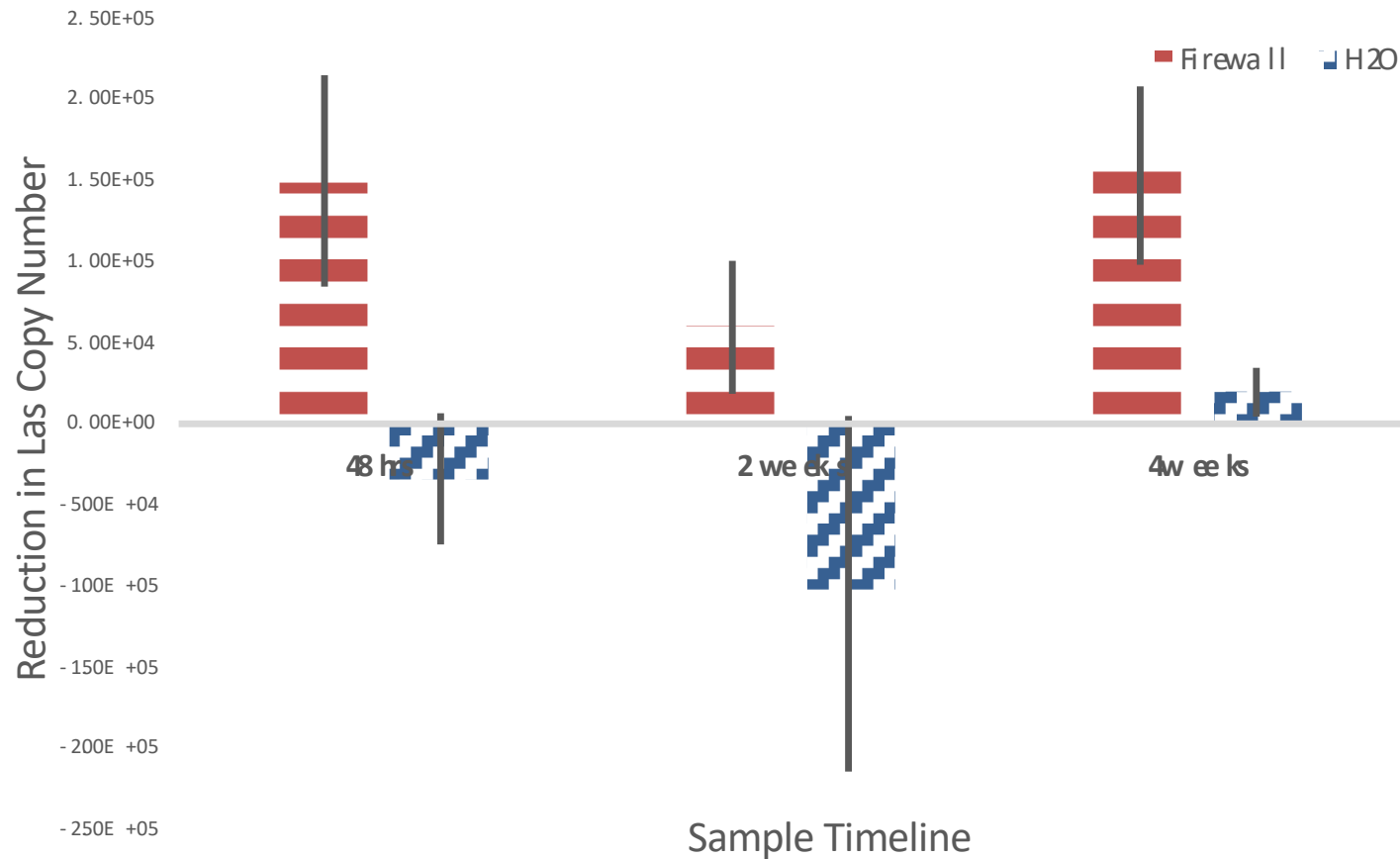
Positive values indicate reductions in CLas titer compared to day 0

Greenhouse Plant Assay: Firewall Vs. Fireline Foliar Spray Efficacy



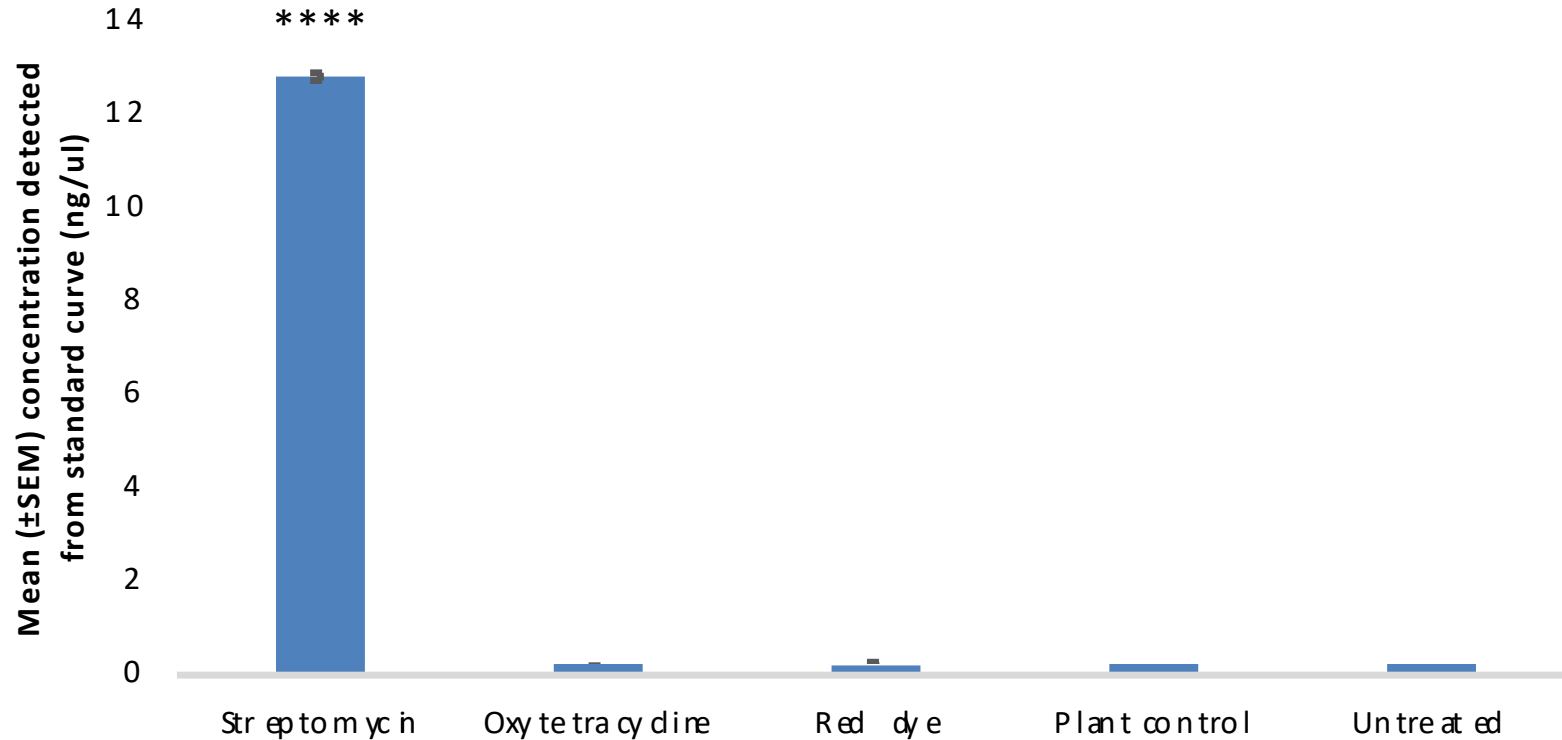
- Positive values indicate reductions in CLas titer compared to day 0 in response to **Firewall** (streptomycin)
- Reduction in CLas after 4 weeks with **Fireline** (oxytetracycline)

Greenhouse Plant Assay: Firewall Soil Drench Efficacy



Positive values indicate reductions in CLas titer compared to day 0 in response to Firewall (streptomycin)

Streptomycin in leaf samples



- Plants treated by soil drench
- Leaves sampled day 6
- Concentrations determined from standard curve in ELISA assay

**** = $P \leq 0.0001$

Do antibiotics reduce ACP feeding?

10 adults (mixed age) released onto a artificial diet ring



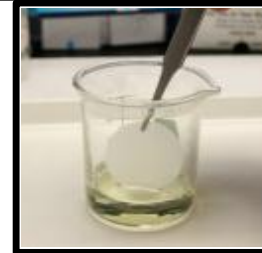
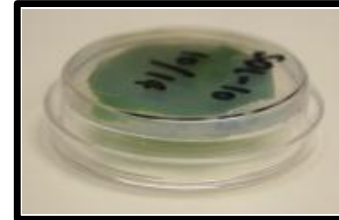
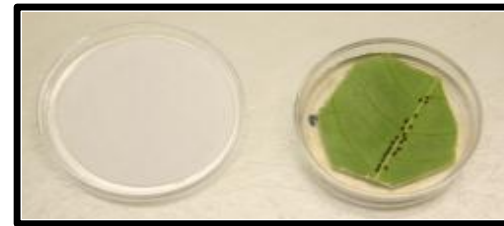
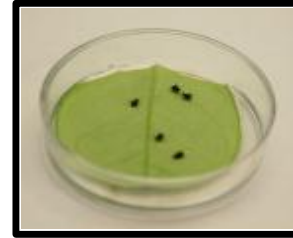
After 3 days, adults removed filter paper kept under the lid



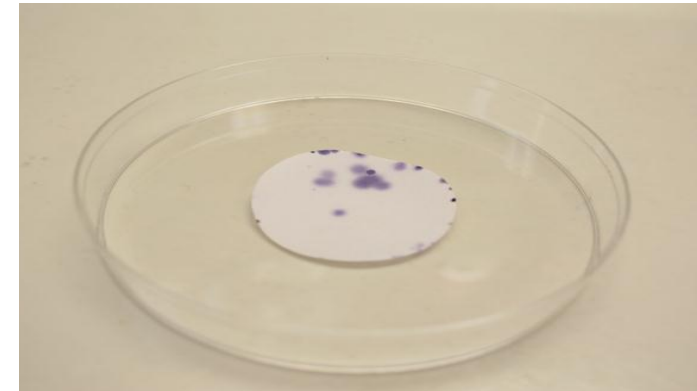
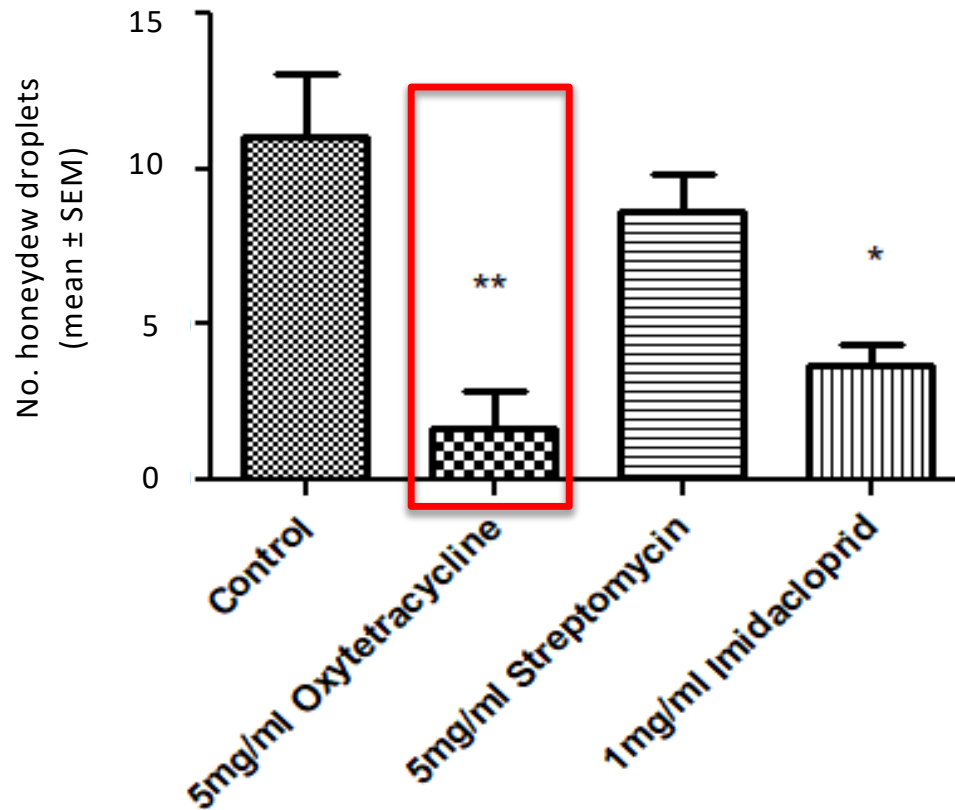
Petri plate inverted to collect honeydew droplets on filter paper



Filter paper dipped in ninhydrin (1%) for 3 min and dried



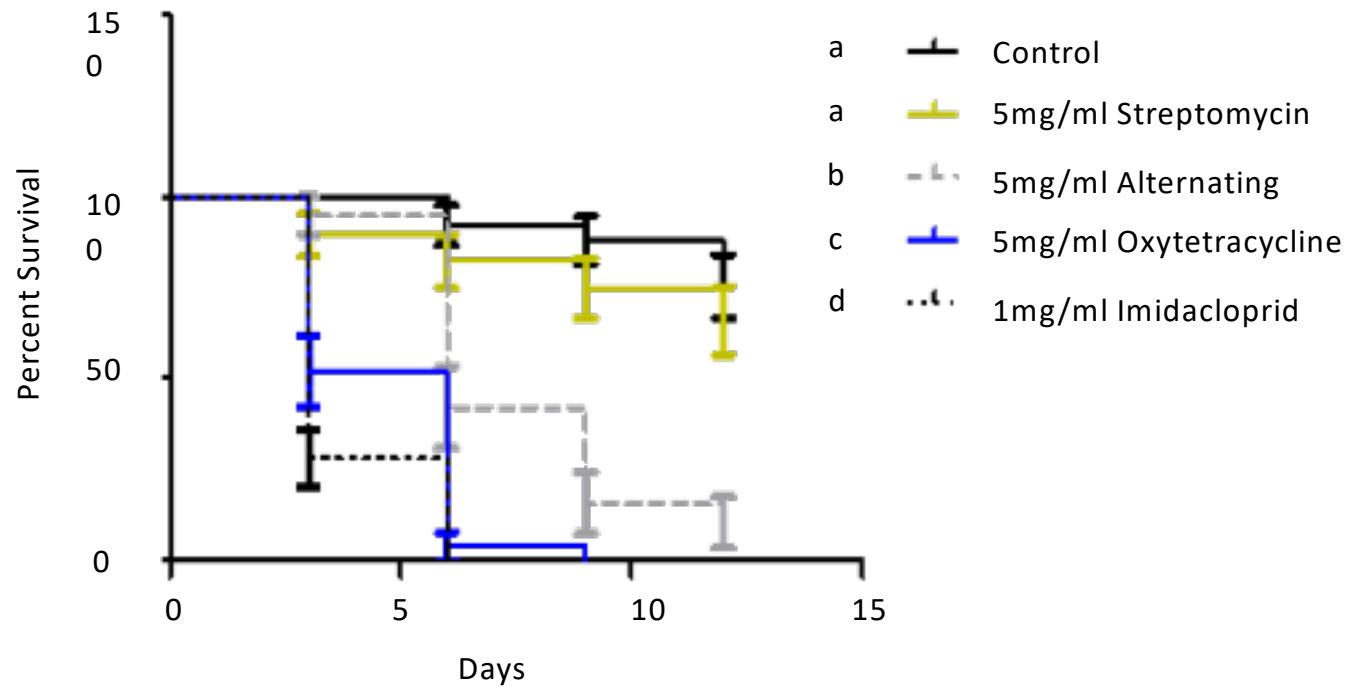
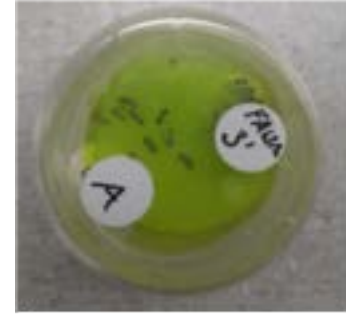
Oxytetracycline inhibits ACP feeding



Purple spots quantified

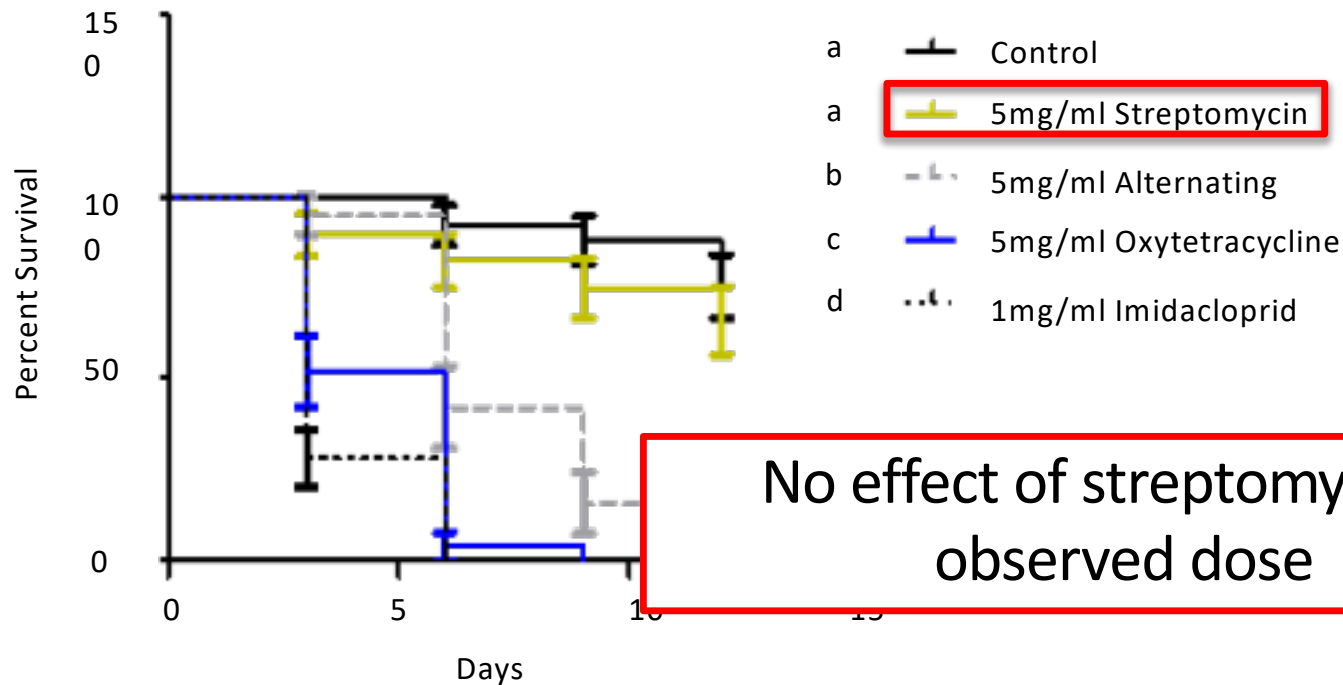
ANOVA: $P = 0.005$, $F = 9.75$, $df = 3, 11$

ACP survival in response to bactericides



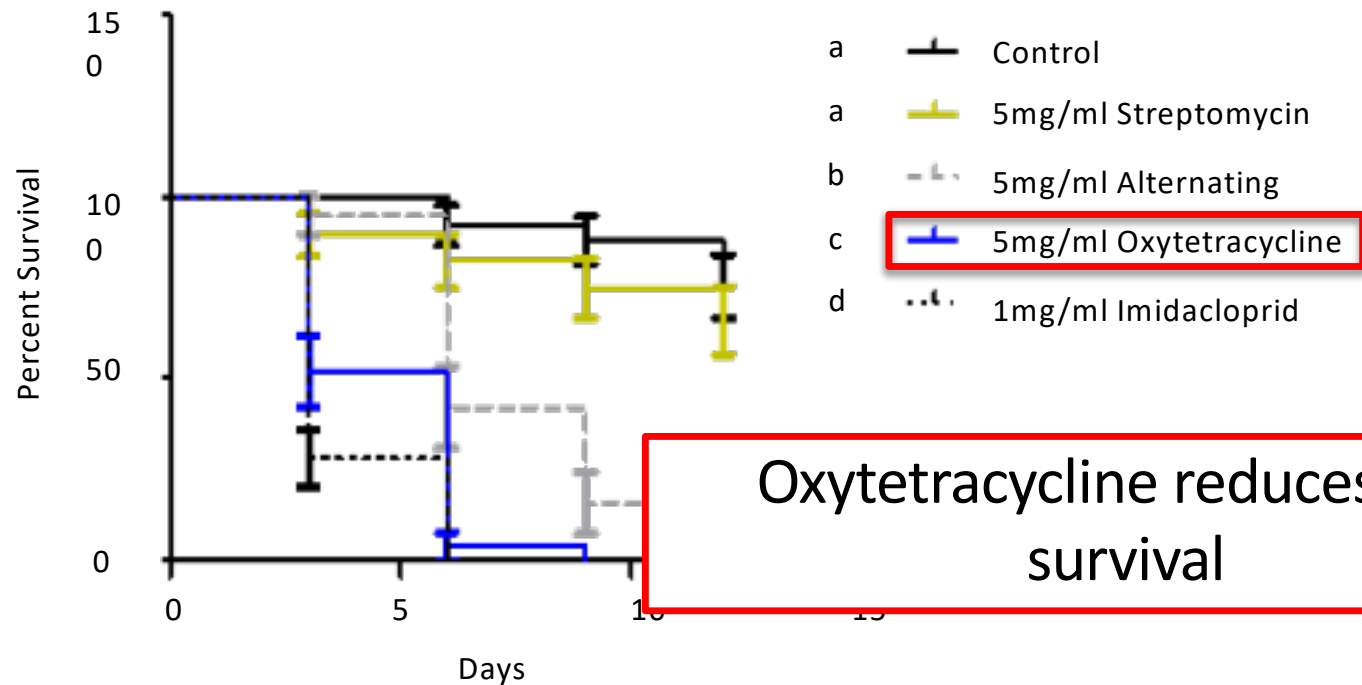
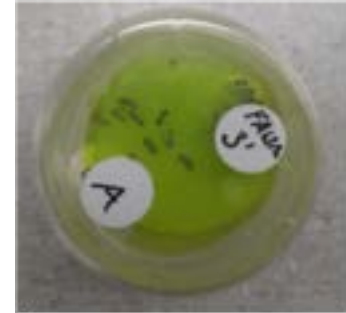
Mantel-Cox: $P = <0.0001$, $F = 5.468$, $df = 4$,

ACP survival in response to bactericides



No effect of streptomycin at observed dose

ACP survival in response to bactericides



Oxytetracycline reduces ACP survival

Antimicrobials for reducing pathogen transmission

Current management of psyllids with broad spectrum insecticides is unsustainable

- High cost
- Physiological resistance

Can we manipulate microorganisms present in the vector and host plant to reduce pathogen transmission?

- Disruption of symbionts for ACP/CLas management

The background of the slide is a grayscale electron micrograph showing numerous bacterial cells. Some cells are rod-shaped, while others are more spherical or irregular. The internal structure of the cells, including what appears to be the cytoplasm and possibly the nucleus or nucleoid region, is visible. The cells are densely packed in some areas and more sparse in others.

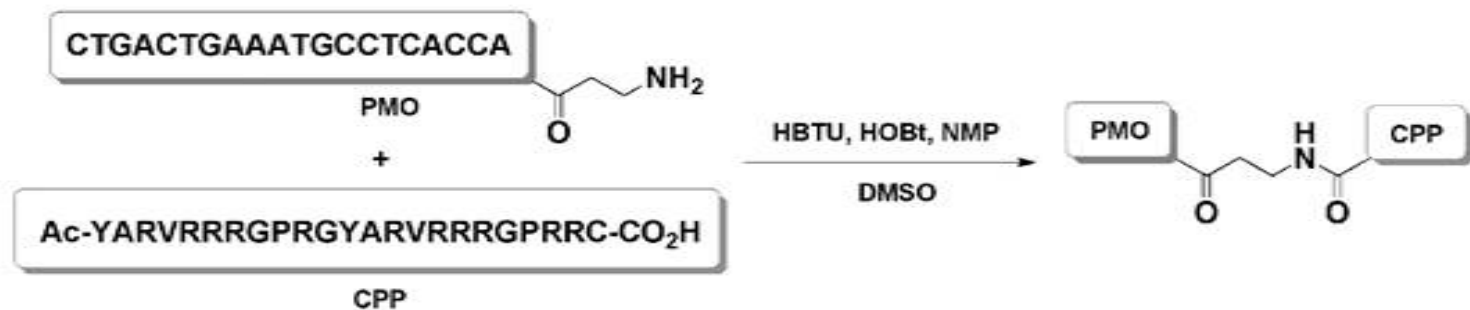
Silencing of bacterial essential genes using FANA/PPMO

— — — — —

. Knock out endosymbiont to reduce transmission/vector fitness

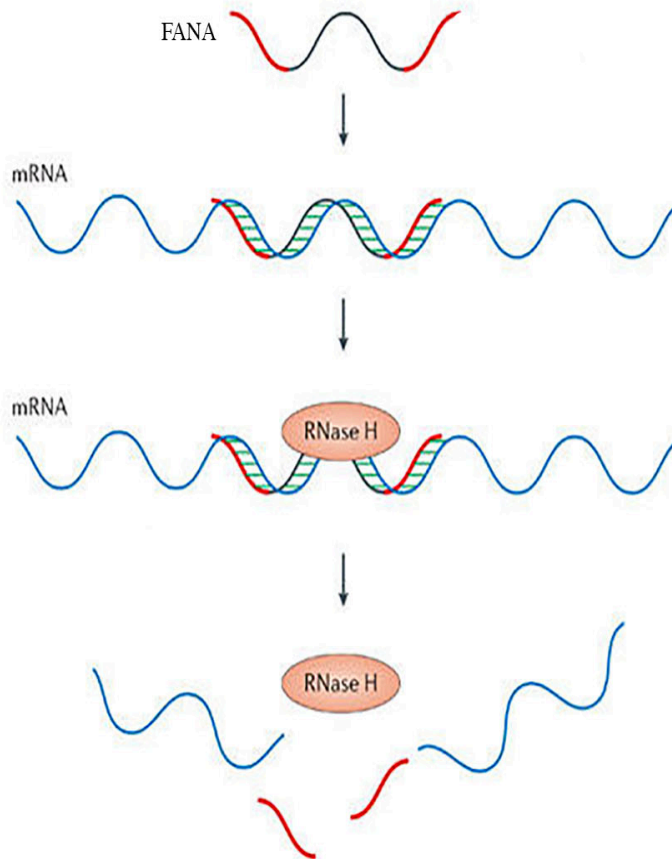
- Development of FANA and morpholino (PMO)-EGS technology targeting CLas and bacterial symbionts in ACP
- Synthetic molecules that mimic DNA and inhibit bacterial gene expression.
- These engineered molecules can be delivered specifically to target bacteria based on gene sequence, avoiding the problems of effecting non-target bacteria.

Synthesis of CPP conjugated PMO



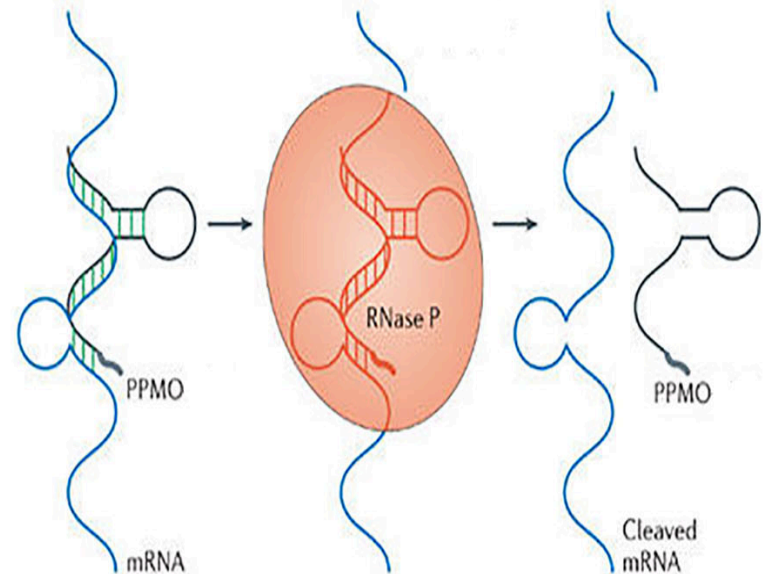
RNA-based bactericidal agents: selectively bind to bacterial mRNA

FANA antisense oligonucleotides (FANA)



- High stability
- Stable hybridization with the target mRNA

Peptide conjugated morpholino (PPMO)

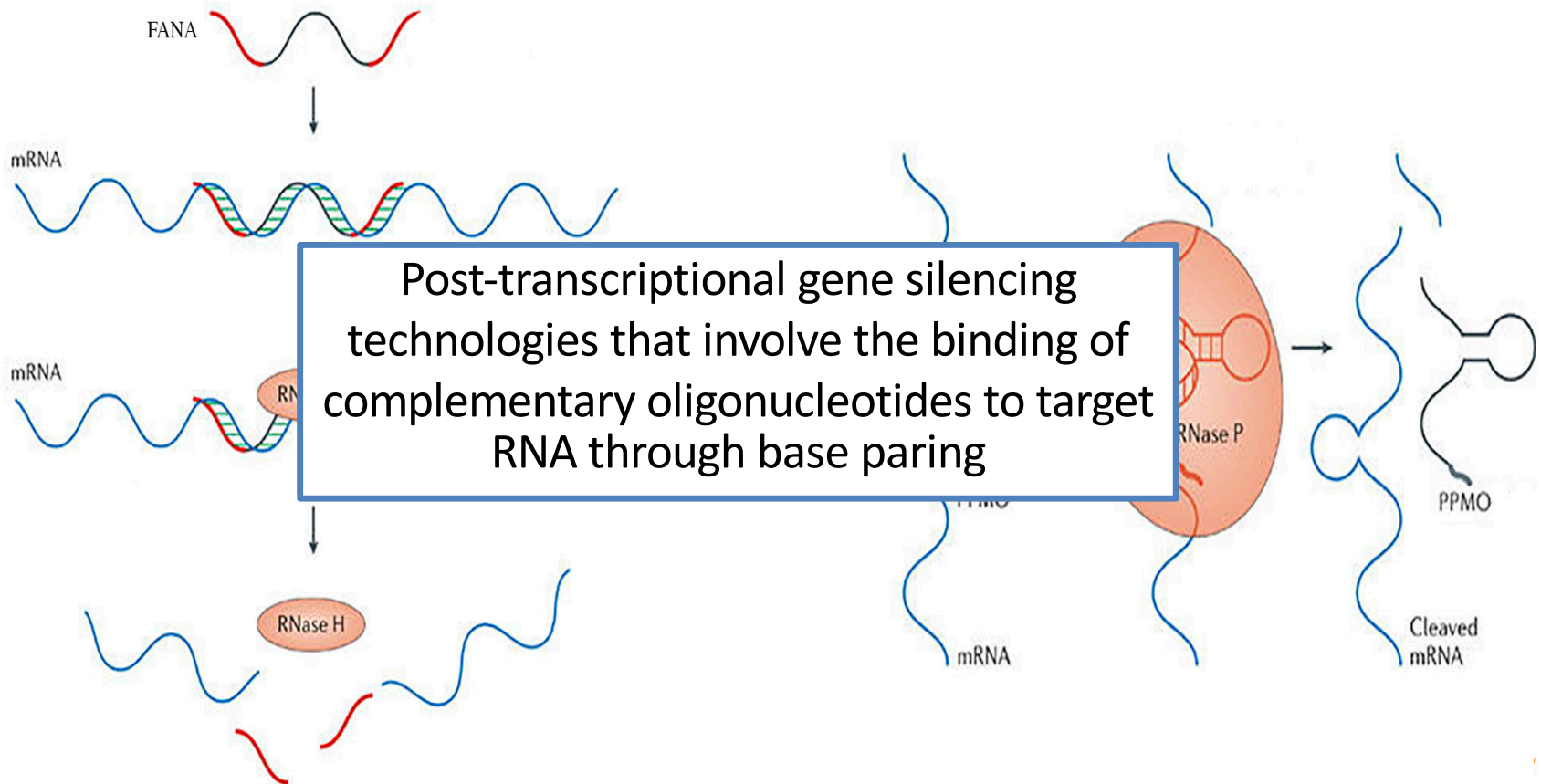


- Endonuclease resistant
- Proven track record of limiting bacterial populations

RNA-based bactericidal agents: selectively bind to bacterial mRNA

FANA antisense oligonucleotides (FANA)

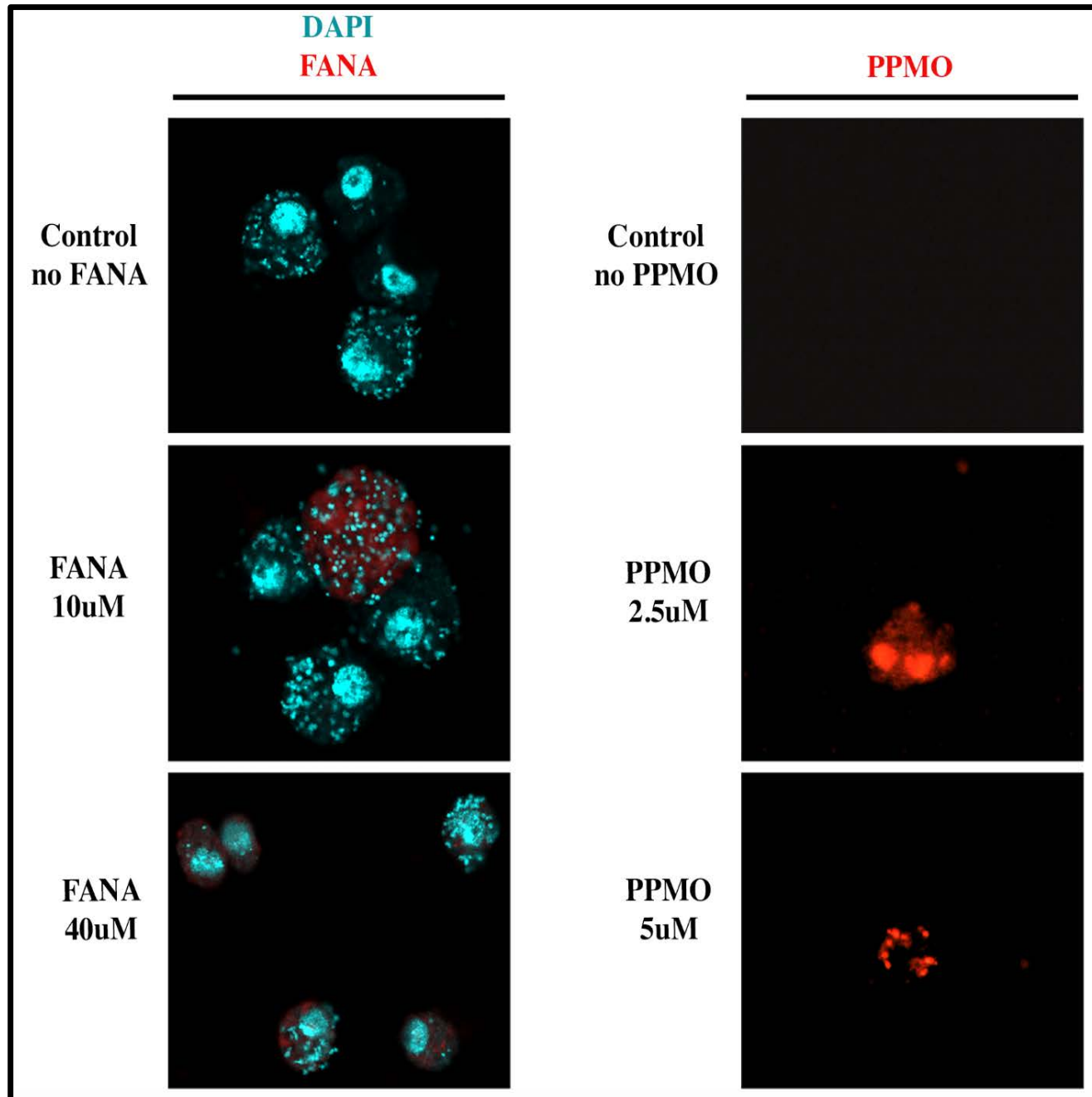
Peptide conjugated morpholino (PPMO)



- High stability
- Stable hybridization with the target mRNA

- Endonuclease resistant
- Proven track record of limiting bacterial populations

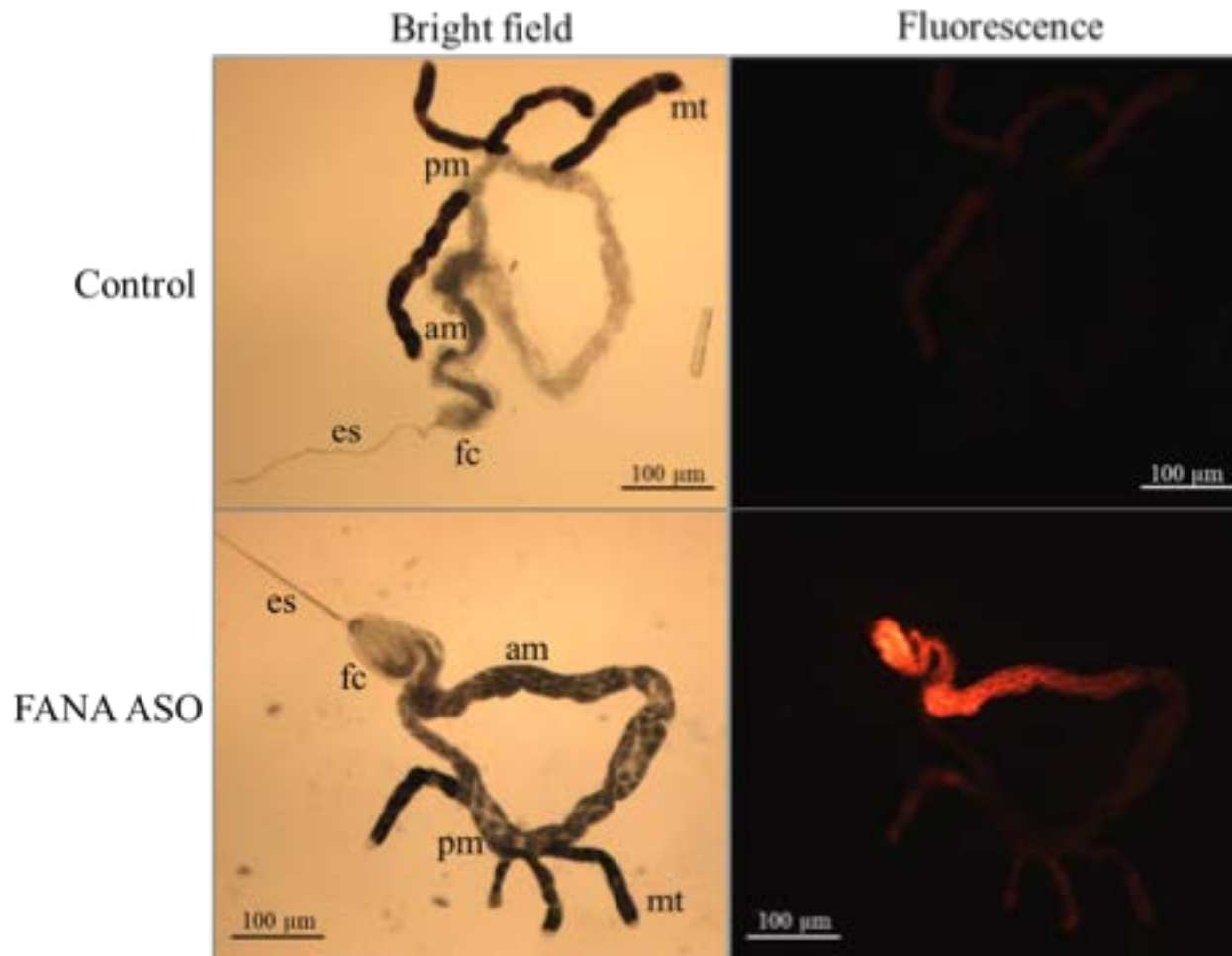
Penetration of FANA and PPMO in insect cells



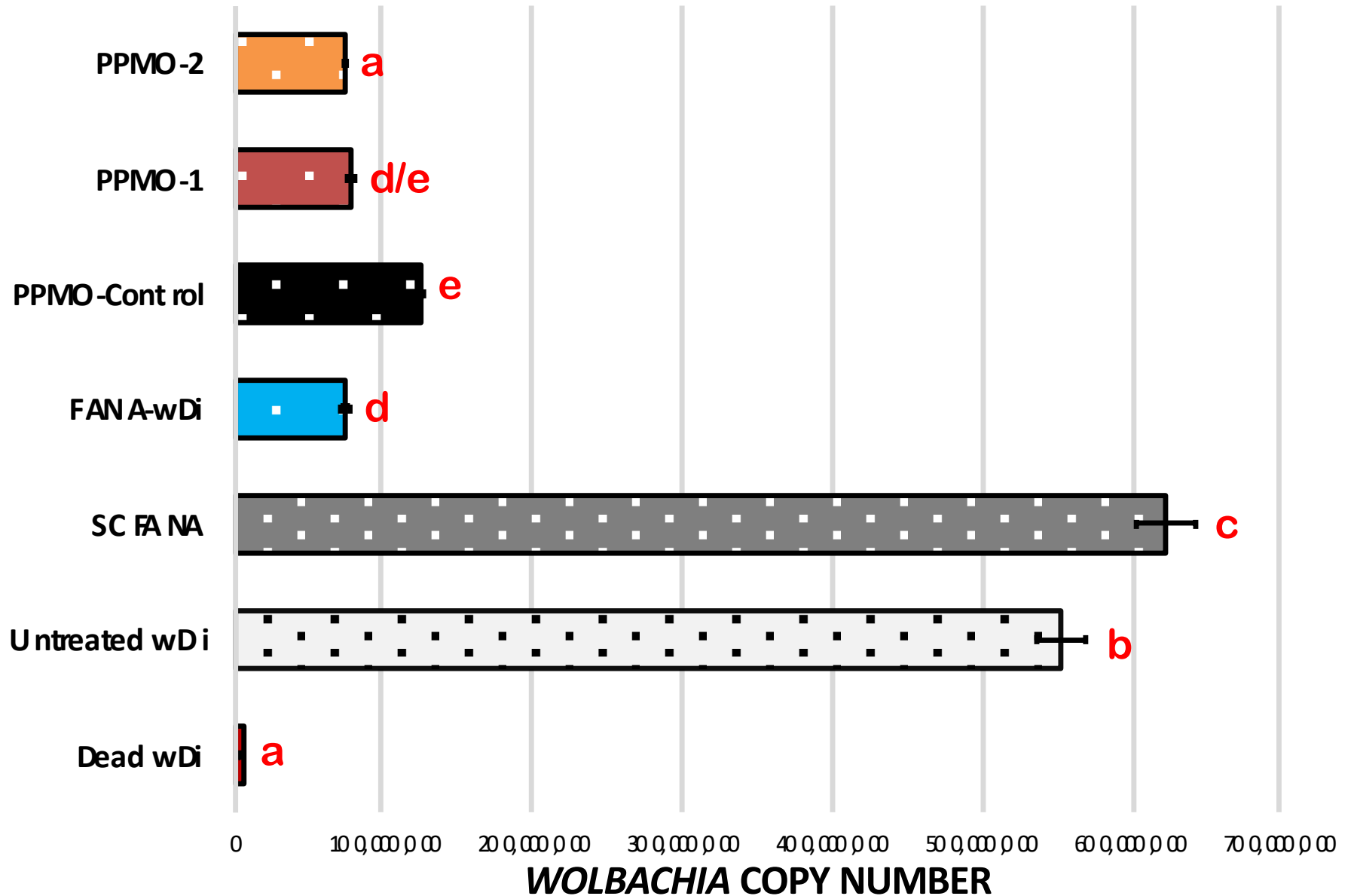
Red indicates
penetration of
oligonucleotides
inside cells

FANA fluorophore:
Alexa647
PPMO fluorophore:
Lissamine

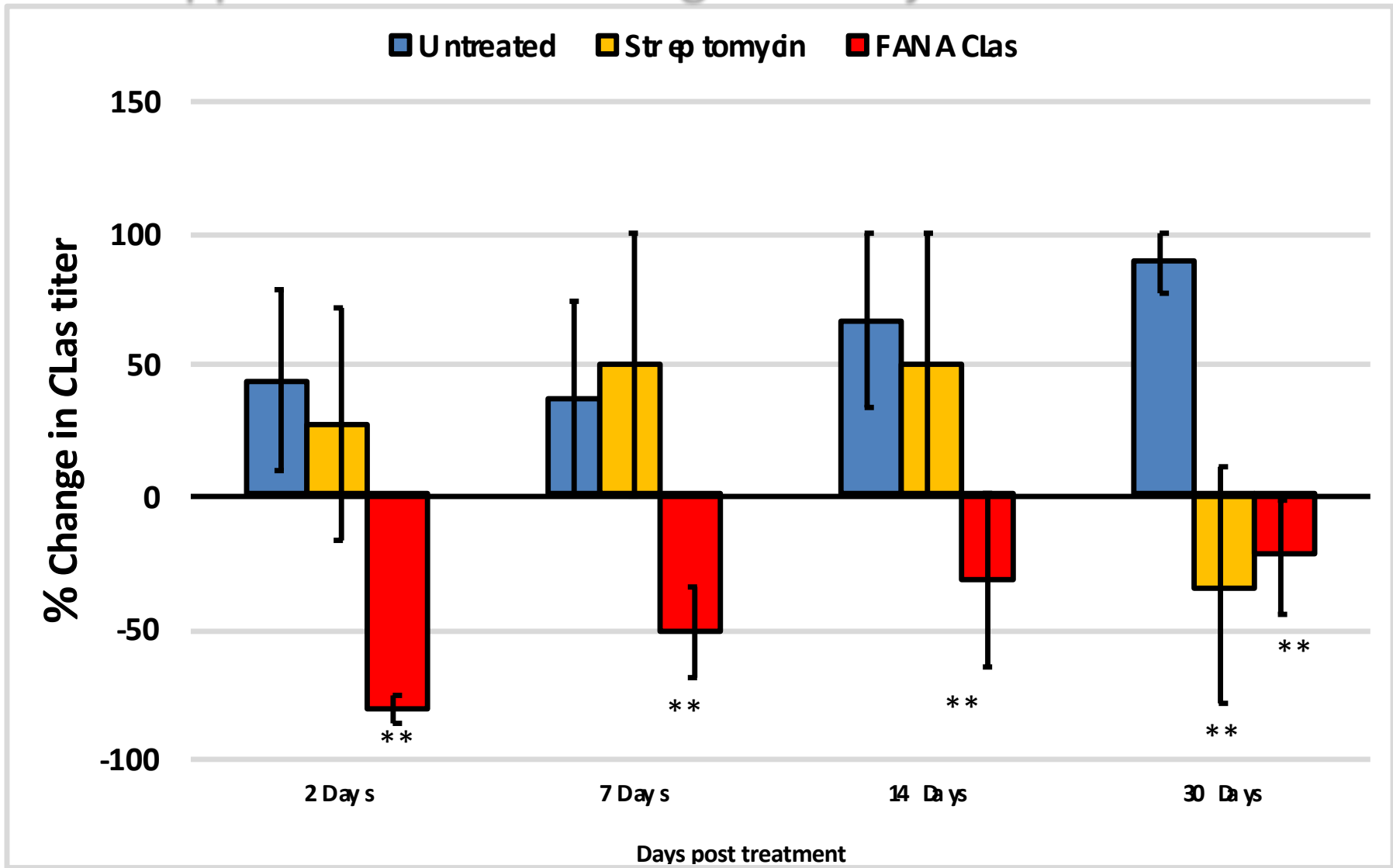
Localization of FANA oligo in the alimentary canal of ACP adults



FANA and PPMOs reduce *Wolbachia* survival in cell culture



FANA applied to citrus through root injection reduces CLas



Negative values represent reduction of Las titer

Summary of antisense efficacy

	FANA	PPMO
Las (Citrus)		
Las (ACP)		
<i>Wolbachia</i> (cell culture)		
Proftella endosymbiont	NA	
ACP Survival		TBD
Transmission	TBD	TBD

Conclusions



- Some reduction of Las titers in response to FireWall and FireLine in greenhouse assays
- Antisense FANAs more effective than streptomycin at reducing Las
- Ingestion of antimicrobials reduces feeding (tetracycline) survival (tetracycline, FANA) of ACP
- Antimicrobial treatments (antibiotics or antisense-RNA technologies) can specifically target Las and microorganisms in ACP

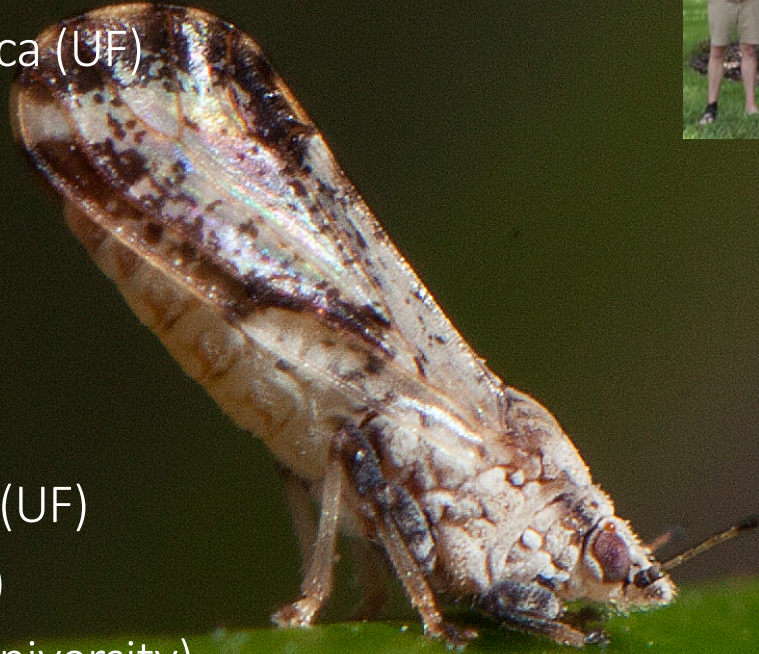
Future Strategies



- Antimicrobials currently available appear to may be effective in reducing ACP populations and CLas transmission
- Targeted antimicrobials likely to be more effective in reducing CLas transmission by ACP
 - Reduces potential for non-target effects
 - Multiple target sites reduce potential for resistance

Thanks!

Andres Sandoval Mojica (UF)
Gustavo Rivas (UF)
Sylvia Bonilla(UF)
Torrence Gill (UF)
Austin McGowan (UF)
Andrew Monalo (UF)
Gennarino Del Bagno (UF)
Wayne Hunter (USDA)
Sidney Altman (Yale University)



Chia-Ching Chu (UF)
Al Handler (USDA)
Evan Braswell (USDA)
Mark Hoffman (NC State)
Calum Russell (UF)
Austin McGowan (UF)
Linhchi Nguyen (UF)
Alex Arp (USDA)



USDA NIFA Project #2016-70016-24782

