

# 2019–2020 Florida Citrus Production Guide: Tristeza Decline<sup>1</sup>

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*Citrus tristeza virus* (CTV) is a major cause of the decline and eventual death of trees on sour orange rootstocks. Initially, affected trees have small leaves and twig dieback. Diseased trees often produce very small fruit. Eventually, large limbs die back and the tree gradually declines. In extreme cases, trees may suffer from quick decline and wilt, dying in a matter of weeks. On sour orange rootstock, some isolates of CTV cause an incompatibility at the bud union, which results in the loss of fibrous roots and reduced ability for water uptake. Bark flaps cut from across the graft union of declining trees often show pitting consisting of small holes (honeycombing) on the inside face of the bark flap from the rootstock side of the union. Quick-decline trees may only have a yellow-brown stain at the bud union and not show the honeycombing. Only trees on sour orange rootstock are affected by tristeza decline. Sweet oranges usually are more affected than grapefruit, whereas lemons on sour orange rootstock are not affected. Furthermore, because huanglongbing (HLB) is endemic in Florida, many trees most likely have a mixed infection with CTV and HLB and may show more profound decline regardless of the rootstock used.

*Citrus tristeza virus* has a wide range of additional isolates of varying symptom severities in different scion-rootstock combinations. Mild isolates have been widespread in

Florida for many years and have been widely disseminated by aphids and in budwood. Such isolates do not usually cause decline of scions on sour orange rootstocks. The incidence of decline has slowed due to the usage of tolerant (non-sour orange) rootstocks such as Swingle citrumelo and Carrizo citrange. However, many of the trees on these tolerant rootstocks are currently infected with CTV decline isolates without showing symptoms. Therefore, there is still a constant high risk for the reappearance of the decline CTV if the sour orange rootstock is used in Florida.

In some countries, CTV isolates that occur also cause stem-pitting in citrus scions (i.e., grapefruit) regardless of the rootstock used. Grooves and pits appear in the wood of the trunk, branches, twigs and rootstocks. Externally, branches may be twisted and ropy, and twigs may become brittle. Trees affected severely by stem-pitting grow poorly and have lower yields and smaller fruits. Limes and grapefruit are most commonly affected by stem-pitting, but sweet oranges also may be affected. Tangerines are generally tolerant of stem-pitting isolates, and show no symptoms. Isolates that cause decline of sweet orange on sour orange also may cause stem-pitting in trees on other rootstocks, but many decline isolates produce no stem-pitting in grapefruit or oranges. Isolates that are closely related to severe stem-pitting isolates found in other countries have

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been found in Florida. Some of these isolates give a degree of sweet orange stem-pitting, but it is difficult to assess whether they cause economic losses in field trees due to widespread presence of HLB.

Virtually all isolates of CTV can be detected by graft inoculation of sensitive biological indicators such as Mexican lime, but more modern and much faster techniques, such as molecular tests including reverse transcriptase (RT)-PCR or ELISA using polyclonal antisera, are nowadays preferred. The monoclonal antibody MCA-13 detects most CTV isolates that cause decline on sour orange rootstocks in Florida, and also reacts with most stem-pitting isolates, but does not react with mild Florida isolates. There are a number of commercial and public research laboratories certified by DPI that can test samples for detection of CTV using RT-PCR, ELISA or tissue-blotting techniques.

Budwood propagated for commercial distribution must be free of MCA-13-reactive CTV. Most stem-pitting isolates are MCA-13-reactive, but not all. Polymerase chain reaction (PCR)-based methods can identify stem-pitting isolates, but the only absolute means to detect stem-pitting isolates is by graft inoculation of grapefruit and sweet orange seedlings and examination of symptoms after 6–15 months (a.k.a. indexing).

*Citrus tristeza virus* is transmitted by aphids. They acquire it within minutes of feeding on an infected plant and transmit it to healthy plants within minutes of picking up the virus. The brown citrus aphid (*Toxoptera citricida*), which first appeared in Florida in 1995, is considered the most efficient vector of the virus. The cotton or melon aphid (*Aphis gossypii*) is a less efficient but still effective vector, whereas the green citrus or spirea aphid (*Aphis spiraecola*) and the black citrus aphid (*Toxoptera aurantii*) are relatively inefficient vectors of CTV in Florida. The establishment of *T. citricida* in Florida is believed to have resulted in a more rapid spread of decline-inducing isolates of tristeza.

## Recommended Practices

1. Budwood propagated for commercial distribution must be free of MCA-13-reactive CTV. Even in the presence of HLB, it is still recommended that farmers only use decline-resistant rootstocks. The key to preventing problems with stem-pitting in Florida is to avoid introduction, propagation, and distribution of stem-pitting isolates.
2. Once tristeza-affected trees on sour orange rootstocks begin to decline, there is no treatment. The diseased trees should be replaced with certified trees on tolerant

rootstocks as the yields of affected trees decline to uneconomical levels. Even though it may be hard to determine if decline results from CTV or HLB, these trees should nonetheless be replaced with a CTV-tolerant rootstock (See the rootstock selection guide)

3. In groves with trees on sour orange rootstock that are currently unaffected by tristeza decline, it may be possible to inarch trees with seedlings of a tolerant rootstock. Inarching is most effective with relatively young groves that are still actively growing, and in order to be effective, the inarches must be well established before trees become infected. Given the high cost of the practice, it is probably advisable only in the case of high-value crops in groves that are well separated from areas with severe strains of tristeza virus. It is more likely to be successful with younger than with older trees.
4. Chemical or biological control of the aphid is unlikely to stop the spread of the virus in commercial groves, since acquisition and transmission of the virus by the aphid usually occurs before the aphid is killed by an insecticide. However, rigorous aphid control in nurseries and on budwood-source trees could reduce infection rates (see ENY-604, *Soft-Bodied Insects*).
5. Cross protection, which is the practice of inoculating trees with mild virus strains to protect them from the effects of severe strains, has been effective in South Africa and Australia in reducing losses in grapefruit due to stem-pitting, and against losses in sweet orange in Brazil. Cross protection against tristeza decline on sour orange rootstock has not yet been developed as an effective control measure.